Breakout groups at EURL-AR Workshop 2023

Dear EURL-AR workshop participants,

At the EURL-AR workshop 2023 (virtual meeting) we will have a session in which NRL representatives will be divided into groups where they will have the opportunity to discuss experiences and challenges observed in EURL-AR EQAS's 2022.

See below the suggestions for discussion items.

In order to obtain the best input for the group discussions, all NRL representatives are encouraged to read and discuss the topics/questions locally prior to attending the workshop, and we welcome you to bring additional observations, challenges or questions into the discussions.

After the groups have had the opportunity to discuss internally, Jette Sejer Kjeldgaard (EURL-AR) will moderate a plenum summary and discussions.

Best regards,

EURL-AR

For discussion:

1. When characterizing an *E.coli* into the categories ESBL, AmpC, carbapenemase producing or 'other phenotypes' (categories defined by EFSA based on phenotypic testing), if an isolate falls into more than one category (e.g., AmpC and 'other phenotypes'), how would you proceed?

2. Do you keep record of the EQA readings (MIC values) to be able to detect systematic trends? How would you react if discovering a systematic, unexpected trend?

3. For reading MIC values, which guidelines do you follow? Which antimicrobials do you consider bacteriostatic? In which cases would you disregard the micropellets?

4. According to your experience, what should be the lowest accepted %ID value for trusting a hit for an antimicrobial resistance gene in the database?

5. Does the prediction of the presence of an antimicrobial resistance gene (even with 100% ID) in a sequence always lead to predicted resistance to the respective antimicrobial?

6. How would you handle a situation where there are hits for more than one variant of the same gene?

7. To identify contamination of sequence data in your routine analysis, which steps would you take?

8. Is long read sequencing technology being used or has it been considered for use in your laboratory? If not, what are the main reasons for this?

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