

Methicillin resistant *Staphylococcus aureus* (MRSA)- Laboratory analysis of dust samples (Baseline methods)



$$f(x+\Delta x) = \sum_{i=0}^{\infty} \frac{(\Delta x)^i}{i!} f^{(i)}(x)$$

$$\Delta \int_a^b \epsilon \Theta^{\sqrt{17}} + \Omega \int \delta e^{i\pi} = \{2.7182818284\}$$

$$\infty = \chi^2 \sum !$$

Detection of MRSA

- Routine susceptibility testing
 - Screening tests (oxacillin, ceftioxin)
 - Confirmation of presence of resistance determinant
 - Molecular detection of *mecA* (PCR)
 - Detection of PBP2a with immunoassays (rapid agglutination tests)
- Selective procedures for isolation



MRSA ST398- Sampling used in baseline study

- Samples: dust samples collected in swabs taken at the farms
 - Pooled sample of 5 soiled swabs of 500cm²
 - Keep in a sterile and identified plastic bag
 - Analysis should start max 13 dys after sample collection



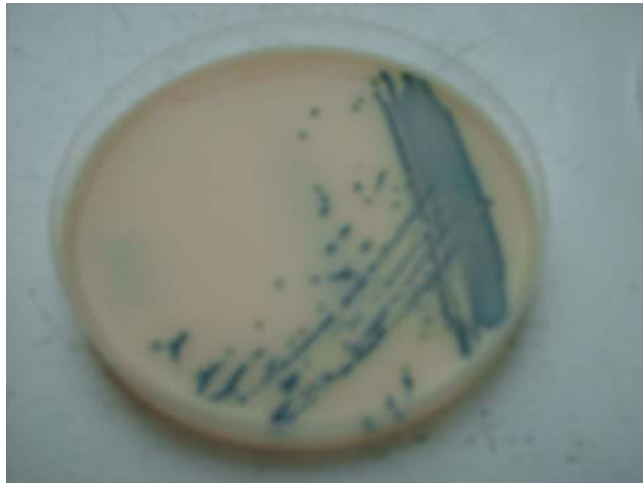
Protocol for selective enrichment of MRSA from dust samples

- Pre-enrichment:
 - The 5 Pooled dust swabs are unpacked and placed in sterile containers
 - 300ml of Mueller Hinton Broth with 6,5% NaCl is added
 - Samples are incubated 16-20h at 37C
- Selective enrichment
 - 1ml of the previous culture is added into
 - Tryptone Soya Broth supplemented with:
 - 3,5 mg/L cefoxitin and 75mg/L aztreonam
 - Incubate 16-20h at 37C



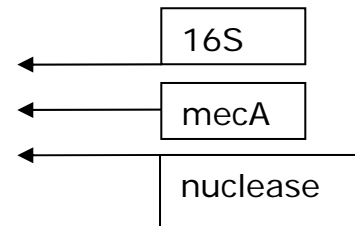
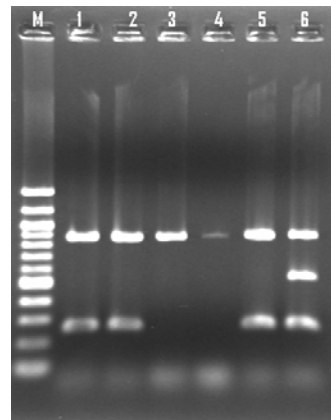
Protocol for isolation of MRSA from dust samples

- Selective plating:
 - Plate 10ul (one blue loop) of the previous selective enrichment onto:
 - Brilliance MRSA Chromogenic agar (Oxoid)
 - Blood agar
 - Incubate 24h at 37C
- Interpretation of results and isolation
 - Check for **blue colonies** on the chromogenic MRSA plate and for typical hemolytic colonies on the blood agar.
 - Isolate 4 blue colonies onto a new blood agar plate
 - Incubate 24h at 37C



Confirmation of MRSA

- Extract DNA of the suspected MRSA isolates
- Confirm the *mecA* gene presence by PCR (multiplex is the recommended)
 - *16S*- confirm the PCR works
 - *mecA* – Confirm methicillin resistance
 - *Nuc*- confirm ID (only positive in *Staphylococcus aureus*)
- Chracterization of strains
 - Typoing
 - *Spa* typing
 - MLST typing (sample)
 - Susceptibility testing (optional)



Typing of MRSA

- Typing of strains
 - PFGE with *Sma*I most frequently (except for ST398 which is not typable with PFGE with *Sma*I) - very discriminatory method
 - Sequence based methods:
 - MLST typing
 - *spa* typing
- Typing of resistance determinant
 - *SCCmec* typing

Rotun *et al.*, 1999.

Questions

